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Supporting Information

Chalcone-Supported Cardiac Mesoderm Induction in Human Pluripotent Stem Cells for Heart Muscle Engineering

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Supporting Information

Supplementary Figures 1-7	Pages 2-8
Supplementary Tables 1-2	Pages 9-10
Legends for Supplementary Videos 1-5	Page 11
Experimental Section	Pages 12-15
Synthesis Schemes	Pages 16-33
References	Page 34

Supplementary Figures

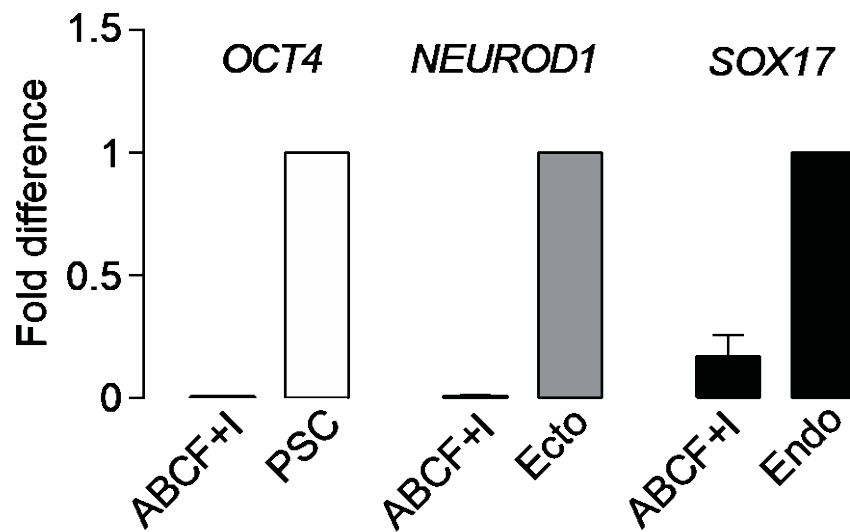


Figure S 1: Negligible transcription of ectodermal and endodermal marker genes in response to the ABCF+I protocol. qPCR screen for residual PSCs (OCT4), ectodermal cells (NEUROD1), and endodermal cells (SOX17; n = 3/group) in ABCF+I-derived heart muscle cells; undifferentiated HES2 and derivatives (ectodermal [Ecto] and endodermal [Endo] induction) served as reference material.

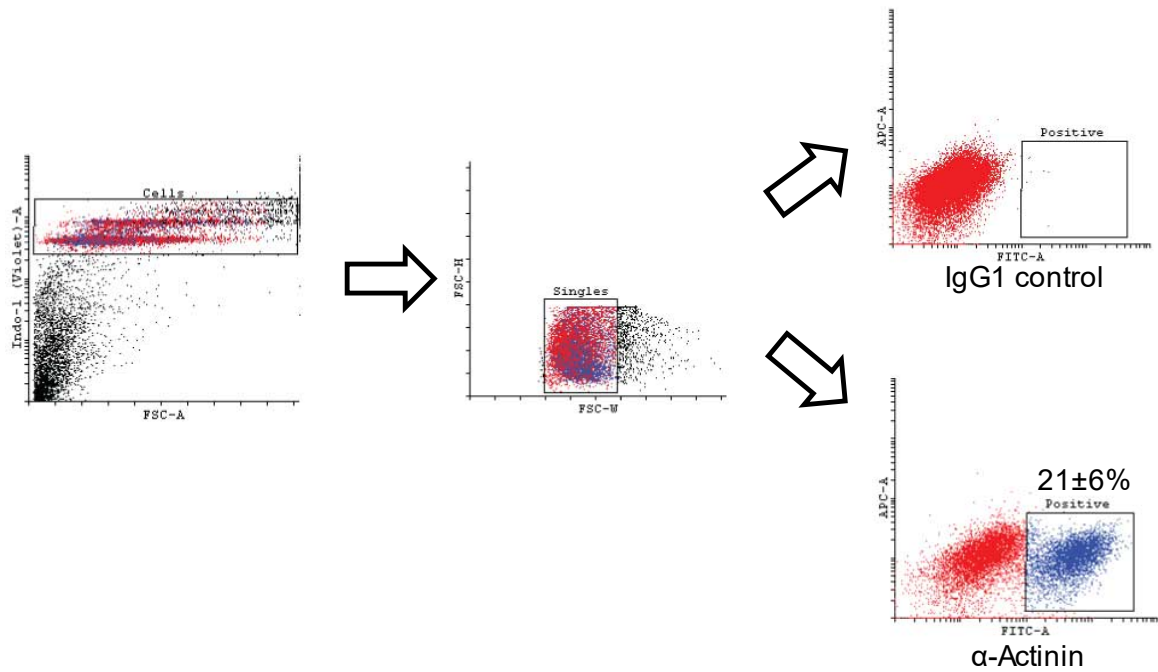


Figure S 2: Gating strategy for the assessment of cardiomyocyte derivation from hPSC by flow cytometry. Intact cells were gated on the DNA label “Hoechst 33342” fluorescence intensity (FI; left panel), then forward scatter width and height (FSC-W; FSC-H) to exclude doublets. Cardiomyocytes were identified by a positive signal after antibody labeling of sarcomeric α -actinin (ACTN2). Representative plots for IgG₁ isotype control and α -actinin on the right (data from $n = 6$ experiments); FSC: forward scatter; FSC-A: FSC-signal area; FSC-H: FSC-signal height; FSC-W: FSC-signal width.

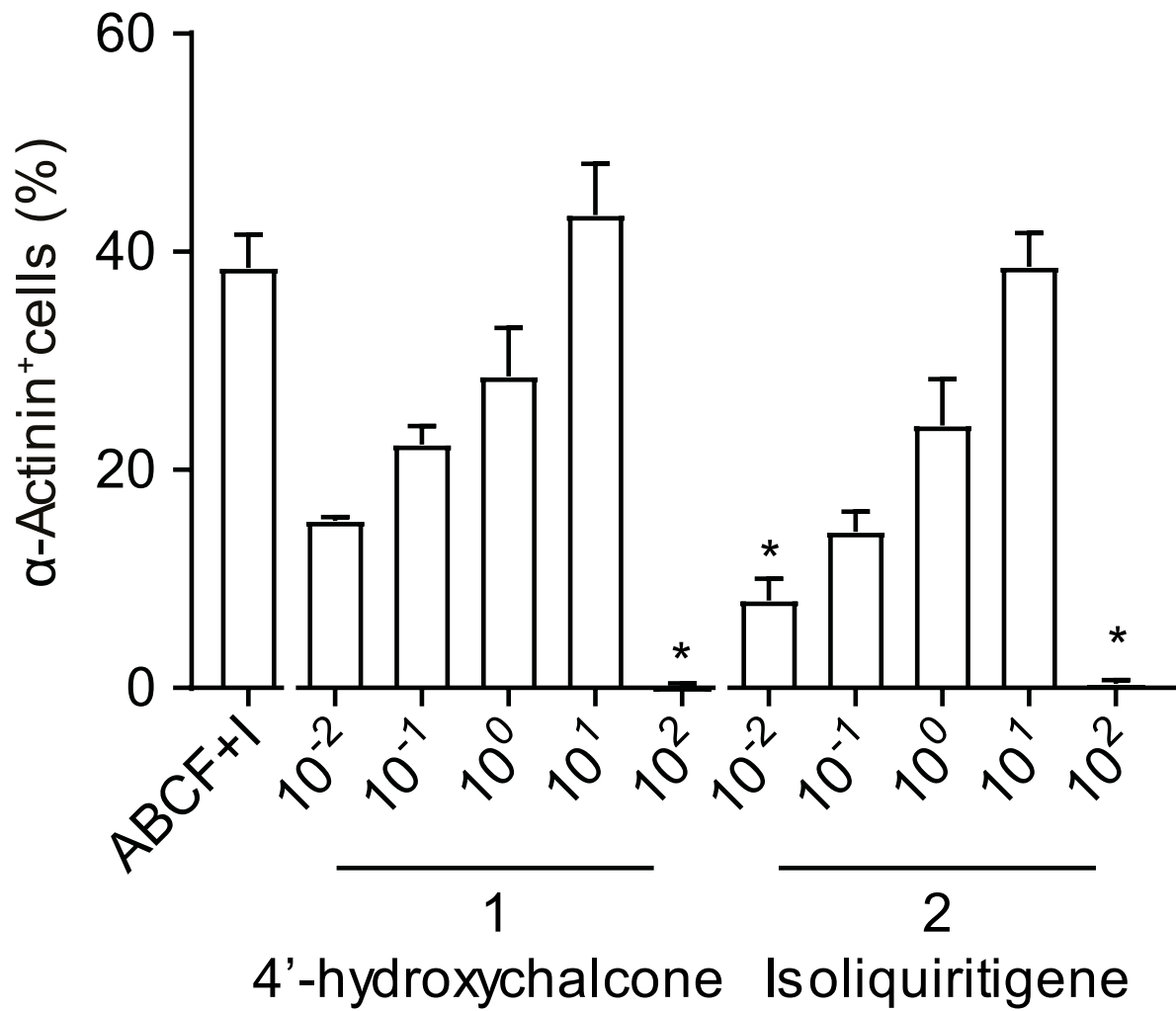


Figure S 3: Chalcone concentration dependent effects on cardiomyogenesis. Flow cytometry analysis of cardiomyocyte content (% α -actinin positive cells on culture day 22) after exposure of hPSCs to the AXCf+I protocol, whereas X refers to the commercially available chalcones 4'-hydroxychalcone (1) and Isoliquiritigene (2) tested at the indicated concentrations (in $\mu\text{mol/L}$; $n = 3-17$ biological replicates/concentration). * $P < 0.05$ vs. ABCF+I by ANOVA with Dunnett's multiple comparison *post-hoc* test.

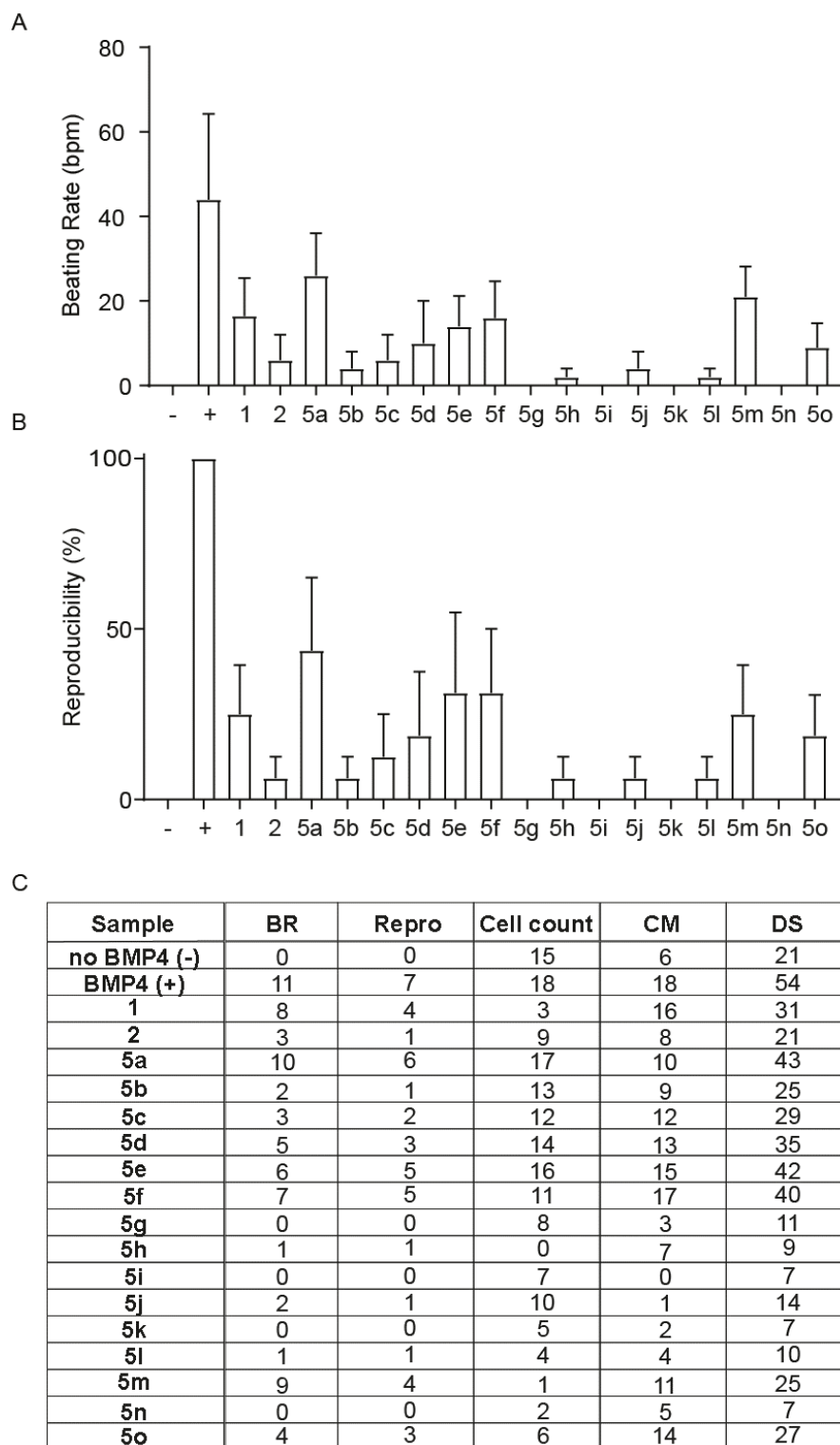


Figure S 4: Primary screening results. Assessment of **(A)** Beating Rate (BR) of the derived cardiomyocytes ($n = 3-4$ /group) and **(B)** Reproducibility (Repro; % of cultures with visibly beating cardiomyocytes - 4 wells analyzed/differentiation run; $n = 4$ differentiation runs/group) at culture day 22 following the screen of 2 commercially available and 15 newly synthesized chalcones. **(C)** Summary of numerical values assigned for each analyzed parameter; CM: cardiomyocytes content in total cell count; DS: differentiation score, summation of the individual scores for each analyzed parameter. - : no BMP4 (ACF+I); + : ABCF+I

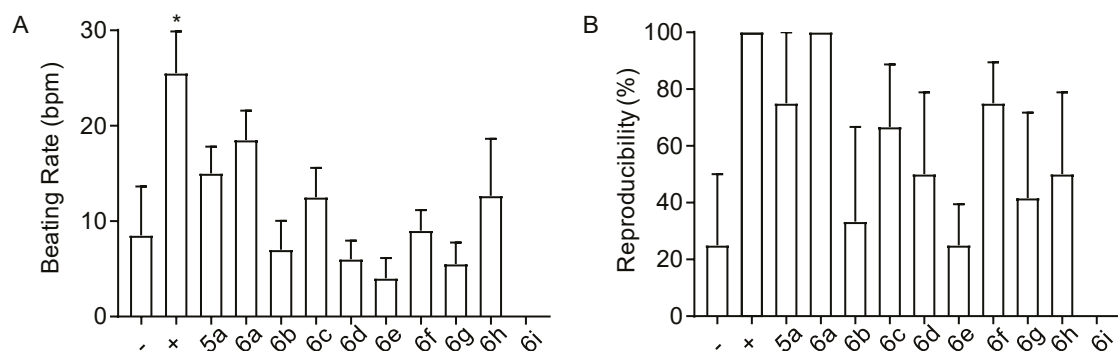


Figure S 5: Screen of 4-fluoro-4'-methoxychalcone (compound 5a) derivatives. Analyses were performed similarly as described for the primary screen of the first round of commercial and newly synthesized chalcones. **(A)** Beating Rate ($n = 12/\text{group}$) and **(B)** Reproducibility (% of cultures with visibly beating cardiomyocytes – 4 wells analyzed/differentiation run; $n = 3$ differentiation runs/group). * $P < 0.05$ vs. no BMP4 (-) by ANOVA with Dunnett's multiple comparison *post-hoc* test. - : no BMP4 (ACF+I); + : ABCF+I

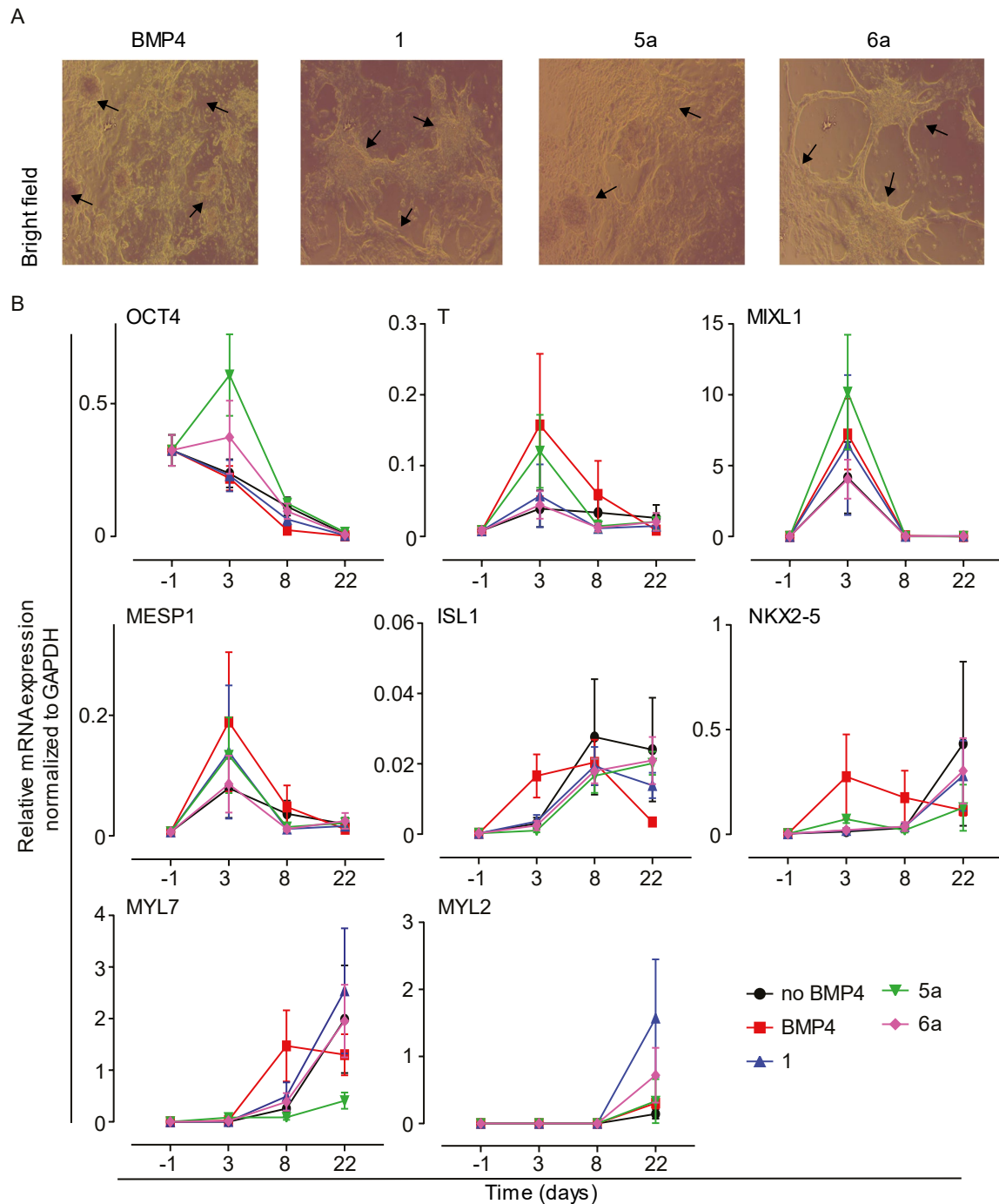


Figure S 6: Morphological and transcript profiling of cardiomyocytes derived by the 4'-hydroxychalcone (1) as well as compound 5a and 6a supplemented AXCF+I protocol. (A) Brightfield images with arrows indicating beating cardiomyocytes clusters; refer also to **Supplementary Videos**. **(B)** Relative gene expression of pluripotent marker *OCT4*, early mesodermal markers *T* and *MIXL1*, cardiac mesodermal marker *MESP1*, early cardiac markers *ISL1* and *NKX2-5* and late cardiac markers *MYL7* and *MYL2* during differentiation using qPCR. All data were normalized to *GAPDH* transcript abundance ($n=3-5$ samples/time point).

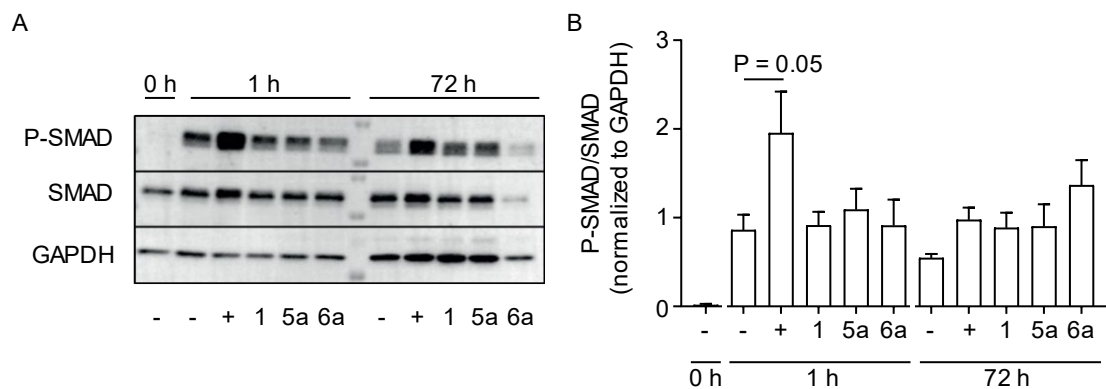


Figure S 7: No evidence for chalcone signaling via the SMAD pathway. (A) Western blot analyses of P-SMAD1/5 and total SMAD1 in cell lysates at 0, 1 (after medium change), and 72 h post-treatment with BMP4 (5 ng/mL), **1**, **5a**, and **6a** (all at 10 μ mol/L). GAPDH was used as loading control (left). (B) Summary of the Western blot data showing relative phosphorylated protein normalized to GAPDH (right; $n=3$ samples/group). - : no BMP4 (ACF+I); + : ABCF+I

Table S1. List of Antibodies used in this study.

(FC – Flow Cytometry, IF – Immunofluorescence, WB – Western Blot)

Antibody/Stain	Company	Cat No.	Target / Use	Dilution
IgG ₁	R&D Systems	MAB002	Control	1:1,000 FC 1:100 IF
α -actinin (mouse IgG)	Sigma	A7811	Cardiomyocyte	1:4,000 FC 1:1,000 IF
α -smooth muscle actin (mouse IgG)	Sigma	A2547	Stromal Cell	1:4,000 FC 1:400 IF
Collagen I (rabbit IgG)	Abcam	Ab34710	Stromal Cell	1:2,000 FC 1:500 IF
Goat anti-mouse Alexafluor 488	Invitrogen	A-11001	Secondary Antibody	1:1,000 FC 1:400 IF
Goat anti-rabbit Alexafluor 546	Invitrogen	A-11010	Secondary Antibody	1:1,000 FC 1:400 IF
Phospho-SMAD1/5 (Rabbit)	Cell Signaling	9516S	Phosphorylated SMAD1/5 (60 kDa)	1:500 WB
SMAD1 (Rabbit)	Cell Signaling	9743S	Total SMAD1 protein (60 kDa)	1:1,000 WB
Anti-mouse-IgG HRP conjugated	Dako	P0161	Secondary Antibody	1:10,000 WB
Anti-rabbit-IgG HRP conjugated	Dako	0448	Secondary Antibody	1:5,000 WB

Table S2. List of Primers used in this study

Gene	F	R	Purpose	Size (bp)	Acc #
OCT4	CAGTGCCCGAAACCCACAC	GGAGACCCAGCAGCCTCAAA	qPCR	161	NM_002701 NM_203289 NM_001173531
T	TCAGCAAAGTCAAGCTCACCA	CCCCAACTCTCACTATGTGGATT	qPCR	102	NM_003181
MIXL1	CCGAGTCCAGGATCCAGGTA	CTCTGACGCCGAGACTTGG	qPCR	58	NM_031944
NKX2-5	ACAACTTCGTGAACTTCGGCG	GTGGACACTCCCGAGTTGCTCT	qPCR	82	NM_001166175.1 NM_001166176.1 NM_004387.3
MESP1	CCACCGTCCCCGCTCCTTCC	CGGTGCTCACAGAGACGGCG	qPCR	130	NM_0018670
ISL1	CGCCTTGCAGAGTGACATAG	GGACTGGCTACCATGCTGTT	qPCR	147	NM_002202.2
MYL7 (MLC2a)	CAGCGGCAAAGGGTGGTGAAC	GGTCCATGGGTGTCAGGGCGAA	qPCR	113	NM_021223.2
MYL2 (MLC2v)	GGCGCAAACCTCAACGTGTT	ACGTTCACTCGCCCAAGGGC	qPCR	149	NM_000432.3
NEUROD1	AGCCACGGATCAATCTTCTC	GCGTGCCTCTAATCATGAAA	qPCR	143	NM_002500.3
SOX17	AGGAAATCCTCAGACTCCTGGGTT	CCCAAACGTGTTCAAGTGGCAGACA	qPCR	111	NM_022454.3
GAPDH	CCTCAAGATCATCAGCAATGCC	ATGTTCTGGAGAGCCCCGC	qPCR	189	NM_002046.3

Video Legends:

Video S1: HES2-cardiomyocytes derived in the absence of BMP4 (ACF+I protocol).

Negative control: No spontaneous beating of cardiomyocytes on culture day 22.

Video S2: HES2-cardiomyocytes derived in the presence of BMP4 (ABCF+I protocol).

Positive control: spontaneous coordinated beating of cardiomyocytes on culture day 22.

Video S3: HES2-cardiomyocytes derived from the AXCF+I protocol whereas X was 4'-hydroxychalcone (compound [cpd] 1). Beating of cardiomyocytes was recorded on culture day 22.

Video S4: HES2-cardiomyocytes derived from the AXCF+I protocol whereas X was 4-fluoro-4'-methoxychalcone (compound [cpd] 5a). Beating of cardiomyocytes was recorded on culture day 22.

Video S5: HES2-cardiomyocytes derived from the AXCF+I protocol whereas X was 4-fluoro-4'-hydroxychalcone (compound [cpd] 6a). Beating of cardiomyocytes was recorded on culture day 22.

Experimental Section

Ethics. The import and use of human embryonic stem cells (HES2-ROSA26-RFP [HES2])^[1]; was approved by the Central Ethics Committee for Stem Cell Research according to the German Stem Cell Act (Az 1710-79-1-4-16).

PSC culture. HES2 were confirmed to be karyotypically normal, mycoplasma free and pluripotent using in vivo teratoma studies, immunofluorescence staining and flow cytometry. Feeder-cell free culture adapted HES2 were seeded at 25,000 cells/cm² on MatrigelTM (1:30 in phosphate-buffered saline [PBS]; BD Biosciences)-coated T75 flasks in TeSR-E8 medium (E8; Stemcell Technologies), supplemented with antibiotics (100 IU/ml penicillin and 100 µg/ml streptomycin; Gibco). After 48 h, HES2 were passaged by dispersion in TrypLE (Gibco) for 3 min at 37 °C and seeded at 50,000 cells/cm² on MatrigelTM coated 24-well plates (culture day - 1).

Standard cardiac differentiation protocol. We applied the ABCF+I cardiac differentiation protocol (**Figure 1A**)^[2] which we introduced previously with robust cardiomyogenesis-inducing activity in all so far tested hPSC lines, including several human embryonic and induced pluripotent stem cell lines. In brief, mesoderm induction was for 3 days with ABCF: Activin A (9 ng/ml), BMP4 (5 ng/ml), CHIR99021 (1 µmol/L) and FGF2 (5 ng/ml) in serum-free (SF) medium (RPMI 1640 supplemented with 2% B27 with insulin, 1 mmol/L sodium pyruvate [all Gibco] and 200 µmol/L L-ascorbic acid 2 phosphate sesquimagnesium salt hydrate [ASC; Sigma] and antibiotics). This was followed by cardiac specification by supplementing SF-medium with IWP4 (5 µmol/L) for 10 days beyond which cultures were maintained in SF medium until day 22.

Screening for a small molecular BMP4 replacement. We tested commercially available (4'-hydroxychalcone [Cat#: CDS003892] and Isoliquiritigene [Cat#: I3766]; both from Sigma-Aldrich) and 24 newly synthesized small molecular weight compounds of the chalcone family (for details see **Synthesis Schemes** below) as BMP4 replacements (AXCF+I protocol). Candidate compounds were ranked according to their cardiomyogenesis inducing activity using a combined differentiation score (DS) derived from an assessment of multiple parameters: (1) reproducibility of cardiomyocyte derivation, (2) cardiomyocyte beating rate, (3) total cell number as well as (4) percentage and (5) total quantity of α -actinin positive cardiomyocytes at culture day 22. More detailed analyses were performed with the most effective compounds as indicated.

Compound Syntheses. A mechanically stirred solution of acetophenone (1 mmol/L) in dry MeOH (3 mL) was treated with a solution of NaOMe (5.4 M in MeOH, 5 mmol/L) at 0 °C and stirring was continued for 1 h at room temperature. Then a solution of aldehyde (1 mmol/L) in MeOH (5 mL) was added dropwise and the reaction mixture was stirred for additional 24 h. The solution was treated with H₂O (20 mL), acidified to pH = 1 with HCl (5 mL of a 10% aq. solution), stirred for 10 min and extracted with EtOAc (4 × 15 mL). The combined organic layers were washed with brine (20 mL), dried over MgSO₄ and concentrated under reduced pressure. Purification by flash column chromatography (EtOAc:PE = 1:9→1:2) yielded the target compounds 5a-o as indicated in **Figure 2B**; synthesis of compounds 5e^[3], j^[4], k^[5], n^[6], and o^[5a, 7] were described previously. A detailed compound structure analyses of all compounds and the synthesis procedures for compounds 6 a-i (summarized also in **Figure 4A**) are listed below.

Chromatography. Thin-layer chromatography was performed on silica gel plates (TLC Silica gel 60 F254; Merck). Column chromatography was performed with silica gel (Geduran[®] Si60, Ø = 32–64 µm; Merck).

NMR spectroscopy. NMR spectra were recorded on Varian “Mercury-300“, “Unity-300“, “Inova-500” and on a “AMX-300” spectrometer from Bruker. Chemical shifts are given in ppm relative to tetramethylsilane (TMS) and coupling constants *J* in Hertz. Solvent signals serve as reference and the chemical shifts converted to the TMS scale (CHCl₃: δ H = 7.24 ppm, δ C = 77.16 ppm). Multiplicities of first order signals are assigned as: s (singlet), sbr (broad singlet), d (doublet), t (triplet), q (quartet), dd (doublet of doublets). Signals of higher order are declared as m (multiplet).

IR spectroscopy. IR spectra were recorded with a FT/IR-4100 spectrometer from JASCO (substances were applied neat on an ATR unit).

UV spectroscopy. UV spectra were recorded on a JASCO V-630 spectrometer.

Mass spectrometry. ESI-MS and ESI-HRMS spectra were recorded on a “Apex IV” spectrometer from Bruker Daltronik.

Cell dissociation for cell counting and flow cytometry. On culture day 22, cells were washed in PBS and dissociated by incubation in dissociation reagent (Accutase [Gibco] supplemented with 0.025% Trypsin [Gibco] and 20 µg/ml DNaseI [Merck]) for 30-45 min. The cells were

pelleted by centrifugation (300 g, 3 min) and fixed with either 4% formaldehyde (FA) for immunofluorescence staining or 70% ice cold ethanol for flow cytometry analysis.

Flow cytometry. Depending on the protein of interest, cells were incubated with primary antibodies (**Table S1**) raised against sarcomeric actinin (Sigma), smooth muscle actin (Sigma) and collagen type I (Abcam) in blocking buffer (containing 5% FBS, 1% bovine serum albumin [BSA], 0.5% Triton X-100 in PBS) for 45 min followed by appropriate secondary antibodies (**Table S1**) in blocking buffer and Hoechst 33342 (10 µg/mL) for 30 min at 4°C. Flow cytometry was performed with a BD LSRII SORP instrument (BD Biosciences). IgG isotype controls (R&D Systems) were used to determine non-specific staining. The gating strategy is displayed in **Figure S2**.

Immunofluorescence staining. Cells were plated on Matrigel™-coated glass coverslips for 24 h in SF-medium before fixation in 4% FA. After blocking for 30 minutes in blocking buffer, primary antibodies (**Table S1**) were added for 90 min followed by secondary antibodies (**Table S1**) and Hoechst 33342 (10 µg/mL) for 60 min at room temperature. Coverslips were mounted onto glass slides (SuperFrost Plus, Thermo Fisher Scientific) using Fluoromount (Dako) and imaged using a Zeiss 710 NLO confocal microscope or a Zeiss AxioImager.M2 fluorescence microscope.

Polymerase chain reaction. Cells were harvested and RNA extraction was performed using Trizol™ following manufacturer's instructions (Ambion). RNA (1 µg) was treated with DNase (Roche) followed by cDNA synthesis using a High Capacity cDNA Reverse Transcription Kit (Applied Biosystems). Quantitative PCR (qPCR) was performed using the Fast SYBR Green Master Mix (Applied Biosystems) in a 384-well format AB7900 HT (Applied Biosystems). Gene expression analyses were performed according to the standard curve method.^[8] *GAPDH* transcript abundance was used for normalization. Primer details are given in **Table S2**.

Immunoblotting. Cells were lysed using CytoBuster reagent (Merck) supplemented with phosphatase- and proteinase-inhibitor cocktail (Roche). Protein concentrations were determined using a Bradford assay. Protein samples were separated by SDS-PAGE and transferred onto polyvinylidene difluoride (PVDF, GE lifesciences) membranes. Membranes were then suspended in blocking buffer containing 5% milk in TRIS-buffered saline with Tween (TBST; 0.02 mol/L TRIS-hydrochloride, 0.17 mol/L NaCl and 0.1% Tween-20 in H₂O) for 1 hour, incubated with primary antibody (**Table S1**; in 5% BSA in TBST) overnight and with secondary antibody at room temperature for 1 hour (**Table S1**). The proteins of interest

were detected using SuperSignal HRP Substrate (Thermo Fisher Scientific) in a ChemiDoc MP Imaging system (Bio-Rad). Signals were normalized to the expression of GAPDH (Zytomed Systems).

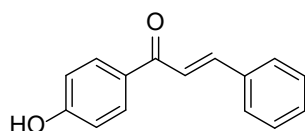
EHM casting. EHM were reconstituted according to a previously described protocol.^[2] Prior to casting and to enhance cardiomyocyte purity, differentiated cells were first subjected to metabolic selection^[9] in RPMI (without Glucose) with 0.1 mmol/L 2-mercaptoethanol, antibiotics (all Gibco) and 2.2 mmol/L sodium lactate (Sigma) for 4 days, followed by a recovery and further maturation phase again in SF medium until culture day 22. This increased cardiomyocyte population to 66±3% for **1** and 24±4% for **5a** (data collected from $n=6-10$ experiments). HES2-derived cardiomyocytes (1.01×10^6 CMs/EHM) and human foreskin fibroblasts (ATCC; 0.44×10^6 cells/EHM) were suspended in a mixture of bovine collagen (LLC Collagen Solutions, 0.4 mg/EHM) and serum-free medium (2x RPMI, 8% B27 without insulin, 200 U/ml penicillin, 200 µg/ml streptomycin). The reconstitution mixture was cast in circular molds (450 µL/EHM) and further cultured in Iscove-Medium containing 4% B27 without insulin, 1% non-essential amino acids, 2 mmol/L glutamine, 200 µmol/L ascorbic acid, 100 U/ml penicillin, and 100 µg/ml streptomycin further supplemented with 100 ng/ml IGF1 (AF-100-11), 10 ng/ml FGF-2 (AF-100-18B), 5 ng/ml VEGF₁₆₅ (AF-100-20) and 5 ng/ml TGF-β1 (AF-100-21C; culture days 0-3 only). All growth factors were purchased from Peprotech.

Isometric force measurements. Force of contraction (FOC, i.e., systolic peak force minus diastolic tension) was measured under isometric conditions in organ baths at 37°C in gassed (5% CO₂/95% O₂) Tyrode's solution (containing in mmol/L: 120 NaCl, 1 MgCl₂, 0.2 CaCl₂, 5.4 KCl, 22.6 NaHCO₃, 4.2 NaH₂PO₄, 5.6 glucose, and 0.56 ascorbate) as previously described.^[2] EHMs were analyzed at 1 Hz with 5 ms square pulses of 200 mA electrical current. EHMs were mechanically stretched at intervals of 125 µm until the maximum twitch force was observed (L_{max}) at 2 mmol/L calcium. Subsequently, maximal inotropic capacity was investigated under increasing extracellular calcium concentrations (0.2-4 mmol/L).

Statistical Analyses. Data are displayed as means with standard error of the mean (SEM) unless stated otherwise. n indicates the number of sample/biological replicates analyzed. Statistical analyses were used for each data set as indicated in the Figure legends using Graph Pad Prism. A P -value of < 0.05 was considered as an indication for significant differences between investigated groups.

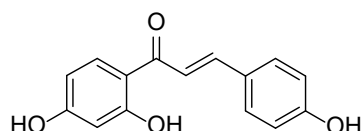
Synthesis Schemes

Scheme 1: Chalcones 1,2, and Synthesis of 5o



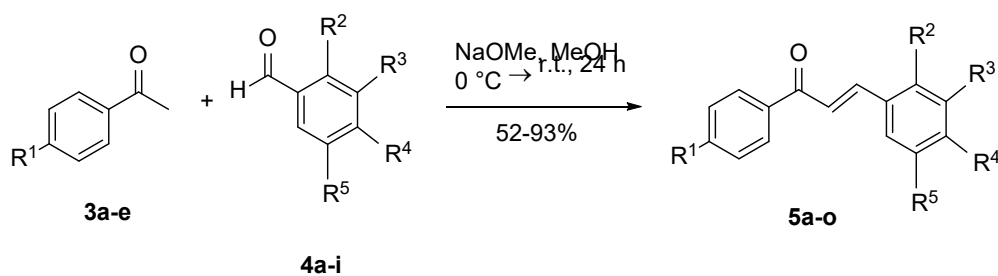
1

4'-Hydroxychalcone (4'HC)
(commercially available)

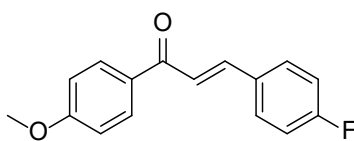


2

Isoquiritigenin (Iso)
(commercially available)



5	a	b	c	d	e	f	g	h	i	j	k	l	m	n	o
R ¹	OMe	OMe	OMe	Me	OH	OH	OMe	NO ₂	F	NO ₂	NO ₂	NO ₂	OMe	F	F
R ²	H	H	H	H	H	H	H	H	H	H	H	H	Br	H	H
R ³	H	H	H	H	H	H	H	H	H	H	H	H	H	OMe	OH
R ⁴	F	OMe	NAC H	F	OMe	NAC H	CO ₂ Me	OMe	CO ₂ Me	F	OH	Me	H	H	H
R ⁵	H	H	H	H	H	H	H	H	H	H	H	H	F	H	H



(E)-3-(4-Fluorophenyl)-1-(4-methoxyphenyl)prop-2-en-1-one (5a):

¹H NMR (300 MHz, CDCl₃): δ 3.87 (s, 3H), 6.93 – 7.01 (m, 2H), 7.00 – 7.12 (m, 2H), 7.46 (d, *J* = 15.6 Hz, 1H), 7.56 – 7.66 (m, 2H), 7.75 (d, *J* = 15.6 Hz, 1H), 7.98 – 8.08 (m, 2H).

¹³C NMR (126 MHz, CDCl₃): δ 55.4 (CH₃), 113.8 (CH), 115.8 (CH), 116.0 (CH), 121.5 (CH), 130.11 (CH), 130.18 (CH), 130.71 (CH), 130.9 (CH), 131.26 (CH), 131.28 (C), 142.5 (CH), 162.8 (C), 164.8 (C), 163.4 (C), 188.3 (C=O).

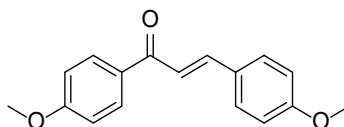
C₁₆H₁₃FO₂ (256.2716)

[M+Na]⁺

calc.: 279.0791

ESI-HRMS

found: 279.0792



(E)-1,3-Bis(4-methoxyphenyl)prop-2-en-1-one (5b):

¹H NMR (500 MHz, CDCl₃): δ 3.85 (s, 3H), 3.88 (s, 3H), 6.91 – 6.95 (m, 2H), 6.96 – 7.00 (m, 2H), 7.43 (d, *J* = 15.6 Hz, 1H), 7.57 – 7.63 (m, 2H), 7.78 (d, *J* = 15.6 Hz, 1H), 8.00 – 8.06 (m, 2H).

¹³C NMR (126 MHz, CDCl₃): δ 55.4 (OCH₃), 113.7 (CH), 114.3 (CH), 119.5 (C), 127.7 (C), 129.9 (CH), 130.5 (CH), 131.2 (C), 143.6 (C), 161.34 (C), 163.1 (C), 188.5 (C=O).

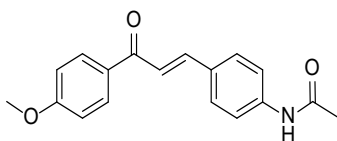
C₁₇H₁₆O₃ (268.3071)

[M+Na]⁺

calc.: 291.0993

ESI-HRMS

found: 291.0992



(E)-N-(4-(3-(4-Methoxyphenyl)-3-oxoprop-1-en-1-yl)phenyl)acetamide (5c):

¹H NMR (500 MHz, CDCl₃): δ 2.20 (s, 3H), 3.89 (s, 3H), 6.95 – 7.01 (m, 2H), 7.48 (d, *J* = 15.6 Hz, 1H), 7.55 (s, 1H), 7.54 – 7.61 (m, 4H), 7.75 (d, *J* = 15.6 Hz, 1H), 8.00 – 8.06 (m, 2H).

¹³C NMR (126 MHz, CDCl₃): δ 24.7 (CH₃), 55.5 (OCH₃), 113.7 (CH), 119.6 (CH), 120.7 (CH), 129.2 (CH), 130.6 (C), 131.0 (C), 139.7 (C), 143.2 (CH), 163.2 (C), 168.2 (CO), 188.5 (CO).

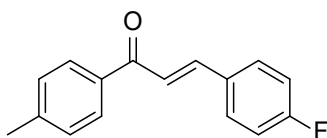
C₁₈H₁₇NO₃ (295.3325)

[M+Na]⁺

calc.: 318.1103

ESI-HRMS

found: 318.1101



(E)-3-(4-Fluorophenyl)-1-(p-tolyl)prop-2-en-1-one (5d):

¹H NMR (500 MHz, CDCl₃): δ 2.44 (s, 3H), 7.06 – 7.15 (m, 2H), 7.31 (dt, *J* = 8.0, 0.8 Hz, 2H), 7.46 (d, *J* = 15.7 Hz, 1H), 7.59 – 7.67 (m, 2H), 7.77 (d, *J* = 15.7 Hz, 1H), 7.89 – 7.98 (m, 2H).

¹³C NMR (126 MHz, CDCl₃): δ 21.7 (CH₃), 115.9 (CH), 116.0 (CH), 121.7 (CH), 128.5 (CH), 129.2 (CH), 130.1 (CH), 131.1 (C), 135.4 (CH), 142.9 (CH), 143.5 (C), 162.8 (C), 164.8 (C), 189.5 (CO).

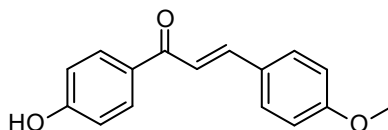
C₁₆H₁₃FO (240.2722)

[M+Na]⁺

calc.: 263.0838

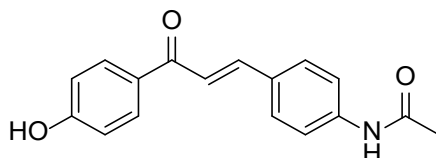
ESI-HRMS

found: 263.0843



(*E*)-3-(4-Methoxyphenyl)-1-(4-hydroxyphenyl)prop-2-en-1-one (5e)^[3]:

C₁₆H₁₄O₃ (254.2806)



(*E*)-*N*-(4-(3-(4-Hydroxyphenyl)-3-oxoprop-1-en-1-yl)phenyl)acetamide (5f):

¹H NMR (300 MHz, d₆-DMSO): δ 2.08 (s, 3H), 6.82 – 6.95 (m, 2H), 7.60 – 7.70 (m, 4H), 7.74 (d, *J* = 8.8 Hz, 2H), 7.92 – 8.08 (m, 2H), 9.95 (s, 1H).

¹³C NMR (126 MHz, d₆-DMSO): δ 24.1 (CH₃), 115.3 (CH), 118.8 (CH), 120.2 (CH), 129.2 (CH), 129.3 (C), 129.4 (C), 130.9 (CH), 141.1 (C), 142.3 (CH), 161.8 (C), 168.5 (CO), 186.9 (CO).

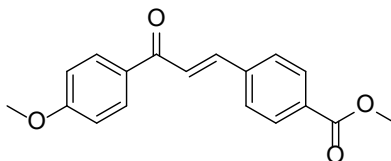
C₁₇H₁₅NO₃ (281.3059)

[M+H]⁺

calc.: 282.1118

ESI-HRMS

found: 282.1125



Methyl (*E*)-4-(3-(4-methoxyphenyl)-3-oxoprop-1-en-1-yl)benzoate (5g):

¹H NMR (400 MHz, CDCl₃): δ 3.90 (s, 3H), 3.94 (s, 3H), 6.96 – 7.04 (m, 2H), 7.62 (d, *J* = 15.7 Hz, 1H), 7.66 – 7.73 (m, 2H), 7.80 (d, *J* = 15.7 Hz, 1H), 8.02 – 8.12 (m, 4H).

¹³C NMR (101 MHz, CDCl₃): δ 52.2 (OCH₃), 55.5 (OCH₃), 113.9 (CH), 123.9 (CH), 128.1 (CH), 130.1 (CH), 130.7 (C), 130.8 (CH), 131.3 (C), 139.3 (C), 142.3 (CH), 163.6 (C), 166.4 (OCO), 188.2 (CO).

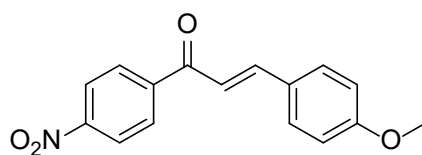
C₁₈H₁₆O₄ (296.3172)

[M+Na]⁺

calc.: 319.0953

ESI-HRMS

found: 319.091



(*E*)-3-(4-Methoxyphenyl)-1-(4-nitrophenyl)prop-2-en-1-one (5h):

¹H NMR (500 MHz, CDCl₃): δ 3.88 (s, 3H), 6.93 – 6.99 (m, 2H), 7.36 (d, *J* = 15.6 Hz, 1H), 7.63 (d, *J* = 15.6 Hz, 1H), 7.79 – 7.86 (m, 1H), 8.10 – 8.16 (m, 2H), 8.32 – 8.38 (m, 2H).

¹³C NMR (126 MHz, CDCl₃): δ 55.5 (OCH₃), 114.5 (CH), 118.9 (CH), 123.7 (CH), 123.9 (CH), 126.9 (CH), 129.2 (CH), 129.3 (C), 130.5 (CH), 143.3 (C), 146.6 (CH), 149.8 (C), 162.1 (C), 188.8 (CO).

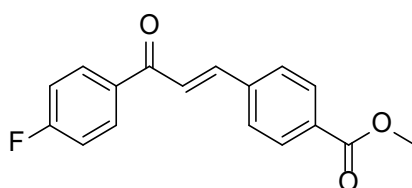
C₁₆H₁₃NO₄ (283.2787)

[M+Na]⁺

calc.: 306.0728

ESI-HRMS

found: 306.0737



Methyl (*E*)-4-(3-(4-fluorophenyl)-3-oxoprop-1-en-1-yl)benzoate (5i):

¹H NMR (300 MHz, CDCl₃): δ 3.91 (s, 3H), 7.08 – 7.22 (m, 2H), 7.55 (d, *J* = 15.7 Hz, 1H), 7.66 (d, *J* = 8.3 Hz, 2H), 7.78 (d, *J* = 15.7 Hz, 1H), 8.05 (ddd, *J* = 8.8, 3.9, 2.1 Hz, 4H).

¹³C NMR (126 MHz, CDCl₃): δ 52.2 (OCH₃), 115.6 (CH), 115.8 (CH), 123.5 (s), 126.5 (C), 128.1 (CH), 129.8 (CH), 130.0 (CH), 131.0 (CH), 131.5 (s), 134.1 (C), 138.8 (CH), 143.2 (CH), 164.6 (C), 166.2 (C), 166.6 (OC=O), 188.2 (C=O).

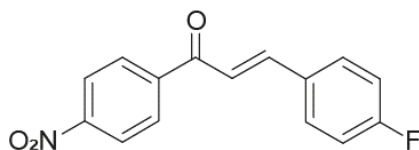
C₁₇H₁₃FO₃ (284.2817)

[M+Na]⁺

calc.: 307.0739

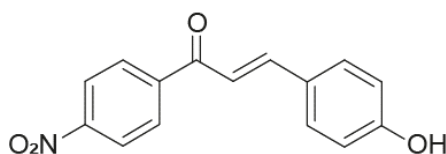
ESI-HRMS

found: 307.0741



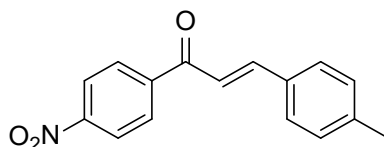
(*E*)-3-(4-Fluorophenyl)-1-(4-nitrophenyl)prop-2-en-1-one (5j)^[4]:

C₁₅H₁₀FNO₃ (271.2432)



(*E*)-3-(4-Hydroxyphenyl)-1-(4-nitrophenyl)prop-2-en-1-one (5k)^[5]:

C₁₅H₁₁NO₄ (269.2521)



(*E*)-1-(4-Nitrophenyl)-3-(p-tolyl)prop-2-en-1-one (5l):

¹H NMR (500 MHz, CDCl₃): δ 2.41 (s, 3H), 7.25 (dt, *J* = 8.4, 0.8 Hz, 2H), 7.44 (d, *J* = 15.7 Hz, 1H), 7.53 – 7.59 (m, 2H), 7.83 (d, *J* = 15.7 Hz, 1H), 8.11 – 8.17 (m, 2H), 8.32 – 8.39 (m, 2H).

¹³C NMR (126 MHz, CDCl₃): δ 21.6 (CH₃), 120.2 (CH), 123.7 (CH), 128.6 (CH), 129.2 (CH), 129.7 (CH), 131.5 (C), 141.8 (C), 143.1 (C), 146.8 (CH), 149.8 (C), 188.9 (CO).

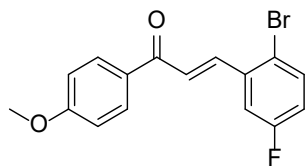
C₁₆H₁₃NO₃ (267.2793)

[M+H]⁺

calc.: 268.0965

ESI-HRMS

found: 268.0968



(*E*)-3-(2-Bromo-5-fluorophenyl)-1-(4-methoxyphenyl)prop-2-en-1-one (5m):

¹H NMR (500 MHz, CDCl₃): δ 3.89 (s, 3H), 6.94 – 7.02 (m, 3H), 7.38 – 7.45 (m, 2H), 7.58 (dd, *J* = 8.8, 5.3 Hz, 1H), 7.98 – 8.07 (m, 3H).

¹³C NMR (126 MHz, CDCl₃): δ 55.5 (OCH₃), 113.8 (CH), 114.2 (CH), 114.4 (CH), 118.3 (CH), 119.8 (C), 125.7 (CH), 130.4 (CH), 130.8 (CH), 134.6 (CH), 136.8 (C), 140.9 (CH), 160.7 (C), 162.7 (C), 163.5 (C), 187.8 (CO).

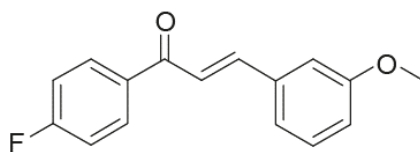
C₁₆H₁₂BrFO₂ (335.1677)

[M+H]⁺

calc.: 335.0081

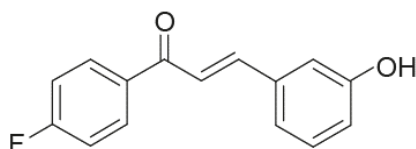
ESI-HRMS

found: 335.0077



(*E*)-3-(3-Methoxyphenyl)-1-(4-fluorophenyl)prop-2-en-1-one (5n)^[6]:

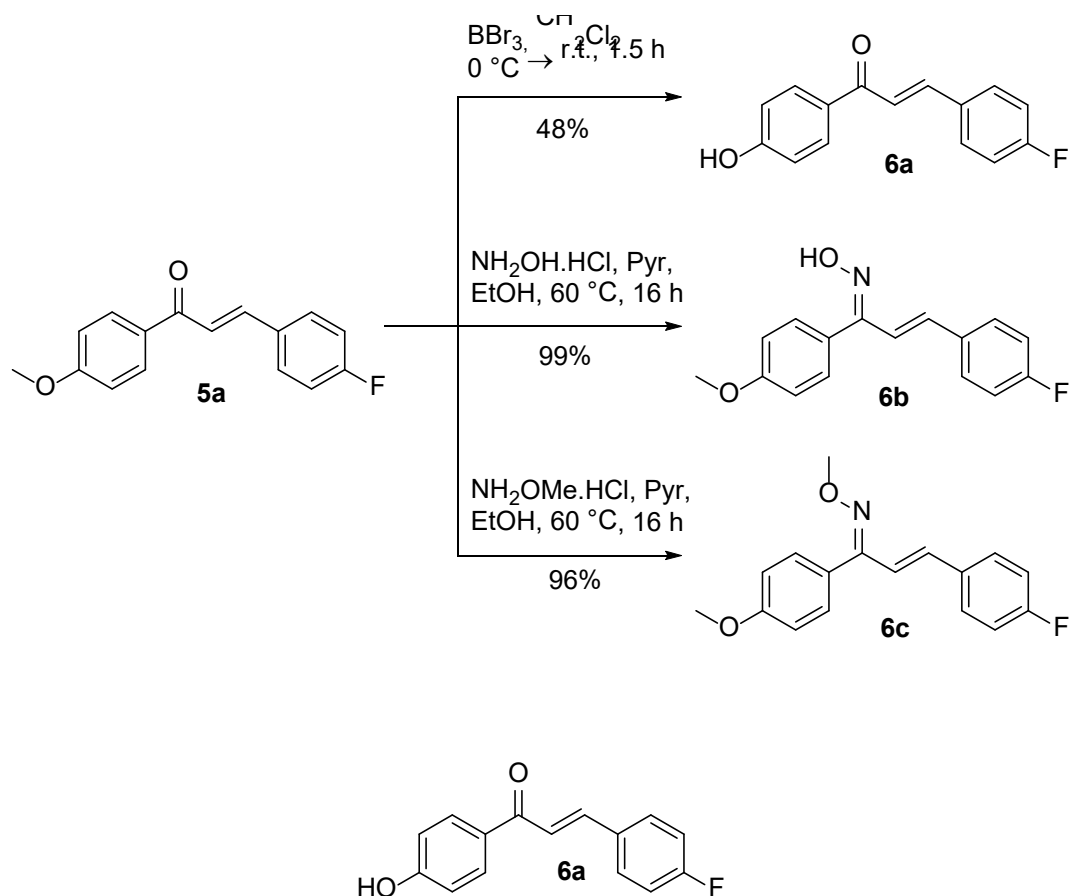
C₁₆H₁₃FO₂ (256.2716)



(*E*)-3-(3-Hydroxyphenyl)-1-(4-fluorophenyl)prop-2-en-1-one (5o)^[5a, 7]:

C₁₅H₁₁FO₂ (242.2450)

Scheme 2: Synthesis of 6a-c



(2E)-3-(4-Fluorophenyl)-1-(4-hydroxyphenyl)prop-2-en-1-one (6a)^[10]: BBr_3 (2.34 mL, 2.34 mmol) in CH_2Cl_2 (3 mL) was added dropwise to a solution of **5a**^[11] (200 mg, 0.78 mmol) in CH_2Cl_2 (3 mL) at 0°C . The reaction mixture was stirred at room temperature for 1.5 h and the reaction quenched with MeOH at 0°C , then the mixture was diluted with water and extracted with CH_2Cl_2 (3 x). The organic phase was dried (Na_2SO_4) and concentrated. The residue was purified by column chromatography on silica gel using 5 – 20% ethyl acetate in petroleum ether to obtain compound **6a** (91 mg, Yield: 48%) as a light orange solid.

^1H NMR (300 MHz, $\text{DMSO}-d_6$) δ 10.36 (s, 1H), 8.06 (d, $J = 8.7$ Hz, 2H), 7.97 – 7.89 (m, 2H), 7.85 (d, $J = 15.6$ Hz, 1H), 7.67 (d, $J = 15.6$ Hz, 1H), 7.27 (t, $J = 8.8$ Hz, 2H), 6.90 (d, $J = 8.7$ Hz, 2H).

^{13}C NMR (126 MHz, $\text{DMSO}-d_6$) δ 186.78, 163.92, 161.95, 141.15, 131.35, 130.90, 130.70, 128.91, 121.92, 115.61, 115.16.

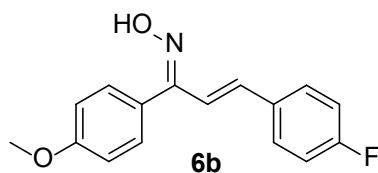
$\text{C}_{15}\text{H}_{11}\text{FO}_2$ (242.2450)

$[\text{M}+\text{Na}]^+$

calc.: 265.0635

ESI-HRMS

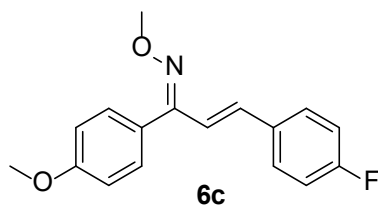
found: 265.0635



***N*-[(1*Z*,2*E*)-3-(4-Fluorophenyl)-1-(4-methoxyphenyl)prop-2-en-1-ylidene]hydroxylamine (**6b**)^[12]:** A mixture of compound **5a** (300 mg, 1.17 mmol), NH₂OH·HCl (244 mg, 3.51 mmol) and pyridine (1 mL) in EtOH (10 mL) was heated at 60°C for 16 h. Then the mixture was concentrated in vacuo, the residue acidified with 1N aq HCl and extracted with ethyl acetate (3 x). The organic phase was dried (Na₂SO₄) and evaporated in vacuo. The obtained crude material was purified by column chromatography on silica gel using 5 – 15% ethyl acetate in petroleum ether to get compound **6b** (314 mg, Yield: 99%) as a white solid.

¹H NMR (300 MHz, CDCl₃) δ 7.57 (d, *J* = 16.5 Hz, 1H), 7.50 – 7.40 (m, 4H), 7.07 – 6.97 (m, 2H), 6.97 – 6.91 (m, 2H), 6.77 (d, *J* = 16.5 Hz, 1H), 3.84 (s, 3H).

C₁₆H₁₄FNO₂ (271.2863)	[M+H] ⁺	calc.: 272.1081
	ESI-HRMS	found: 272.1081

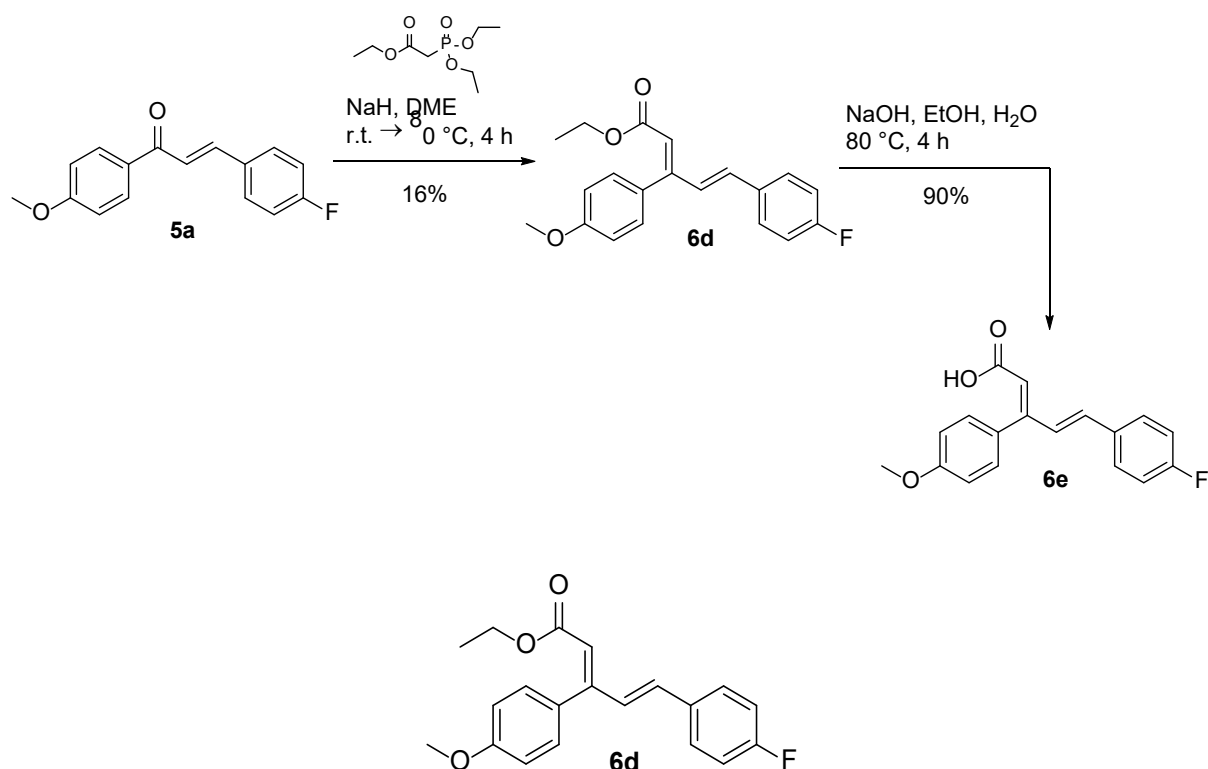


(1*Z*,2*E*)-3-(4-Fluorophenyl)-*N*-methoxy-1-(4-methoxyphenyl)prop-2-en-1-imine (6c**):** A mixture of compound **5a** (300 mg, 1.17 mmol), MeONH₂·HCl (293 mg, 3.51 mmol) and pyridine (1 mL) in EtOH (10 mL) was heated at 60°C for 16 h. The mixture was concentrated in vacuo, the obtained residue acidified with 1N aq HCl and extracted with ethyl acetate (3 x). The organic phase was dried (Na₂SO₄) and evaporated in vacuo. The obtained crude material was purified by column chromatography on silica gel using 5 – 15% ethyl acetate in pet ether to get compound **6c** (321 mg, Yield: 96%) as a white solid.

¹H NMR (300 MHz, CDCl₃) δ 7.53 – 7.40 (m, 4H), 7.30 – 7.22 (m, 1H), 7.06 – 6.89 (m, 4H), 6.73 (d, *J* = 16.6 Hz, 1H), 4.04 (s, 3H), 3.83 (s, 3H).

C₁₇H₁₆FNO₂ (285.3128)	[M+H] ⁺	calc.: 286.1239
	ESI-HRMS	found: 286.1238

Scheme 3: Synthesis of 6d and 6e



Ethyl (2*Z*,4*E*)-5-(4-fluorophenyl)-3-(4-methoxyphenyl)penta-2,4-dienoate (6d): A solution of triethylphosphonoacetate (874 mg, 3.9 mmol) in dimethoxyethane (1 mL) and a solution of compound **5a** (1 g, 3.9 mmol) in dimethoxyethane (7 mL) were successively added to a suspension of 60% NaH (156 mg, 3.9 mmol) in dimethoxyethane (7 mL) at room temperature. The reaction mixture was heated at 80°C for 4 h, then cooled to room temperature, diluted with water (100 mL), and extracted with CH₂Cl₂ (3 x). The organic phase was dried (Na₂SO₄) and evaporated in vacuo. The obtained crude material was purified by column chromatography on silica gel using 5 – 10% ethyl acetate in petroleum ether to afford compound **6d** (206 mg, Yield: 16%) as a colourless thick liquid.

¹H NMR (300 MHz, CDCl₃) δ 8.40 (dt, *J* = 16.2, 0.8 Hz, 1H), 7.50 – 7.38 (m, 2H), 7.30 (d, *J* = 8.7 Hz, 2H), 6.99 (t, *J* = 8.7 Hz, 2H), 6.91 (d, *J* = 8.7 Hz, 2H), 6.59 (d, *J* = 16.2 Hz, 1H), 5.78 (d, *J* = 0.9 Hz, 1H), 4.22 (q, *J* = 7.1 Hz, 2H), 3.84 (s, 3H), 1.32 (t, *J* = 7.1 Hz, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 166.35, 163.84, 161.86, 159.92, 155.27, 137.90, 132.87, 132.12, 130.19, 128.96, 126.00, 117.03, 115.59, 113.63, 59.96, 55.33, 14.45.

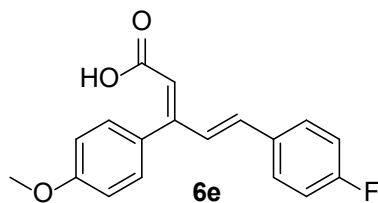
C₂₀H₁₉FO₃ (326.3615)

[M+Na]⁺

calc.: 349.1205

ESI-HRMS

found: 349.1210



(2Z,4E)-5-(4-Fluorophenyl)-3-(4-methoxyphenyl)penta-2,4-dienoic acid (6e): A mixture of compound **6d** (120 mg, 0.37 mmol) and NaOH (147 mg, 3.67 mmol) in EtOH (6 mL) and water (1.5 mL) was heated at 80°C for 4 h. The reaction mixture was cooled to room temperature, acidified with 2N HCl, extracted with ethyl acetate (3 x). The organic phase was dried (Na₂SO₄) and concentrated under vacuum. The resulting solid was washed with pet ether to get the product **6e** (97 mg, Yield: 90%) as a white solid.

¹H NMR (300 MHz, DMSO-*d*₆) δ 12.27 (s, 1H), 8.29 (d, *J* = 16.3 Hz, 1H), 7.53 (d, *J* = 8.7, 2H), 7.33 (d, *J* = 8.6 Hz, 2H), 7.21 (t, *J* = 8.9 Hz, 2H), 7.01 (d, *J* = 8.7 Hz, 2H), 6.59 (d, *J* = 16.3 Hz, 1H), 5.72 (s, 1H), 3.81 (s, 3H).

¹³C NMR (126 MHz, DMSO-*d*₆) δ 166.68, 163.01, 161.05, 159.48, 153.50, 136.40, 132.69, 131.28, 129.83, 128.85, 125.83, 117.88, 115.59, 113.74, 55.07.

C₁₈H₁₅FO₃ (298.3083)

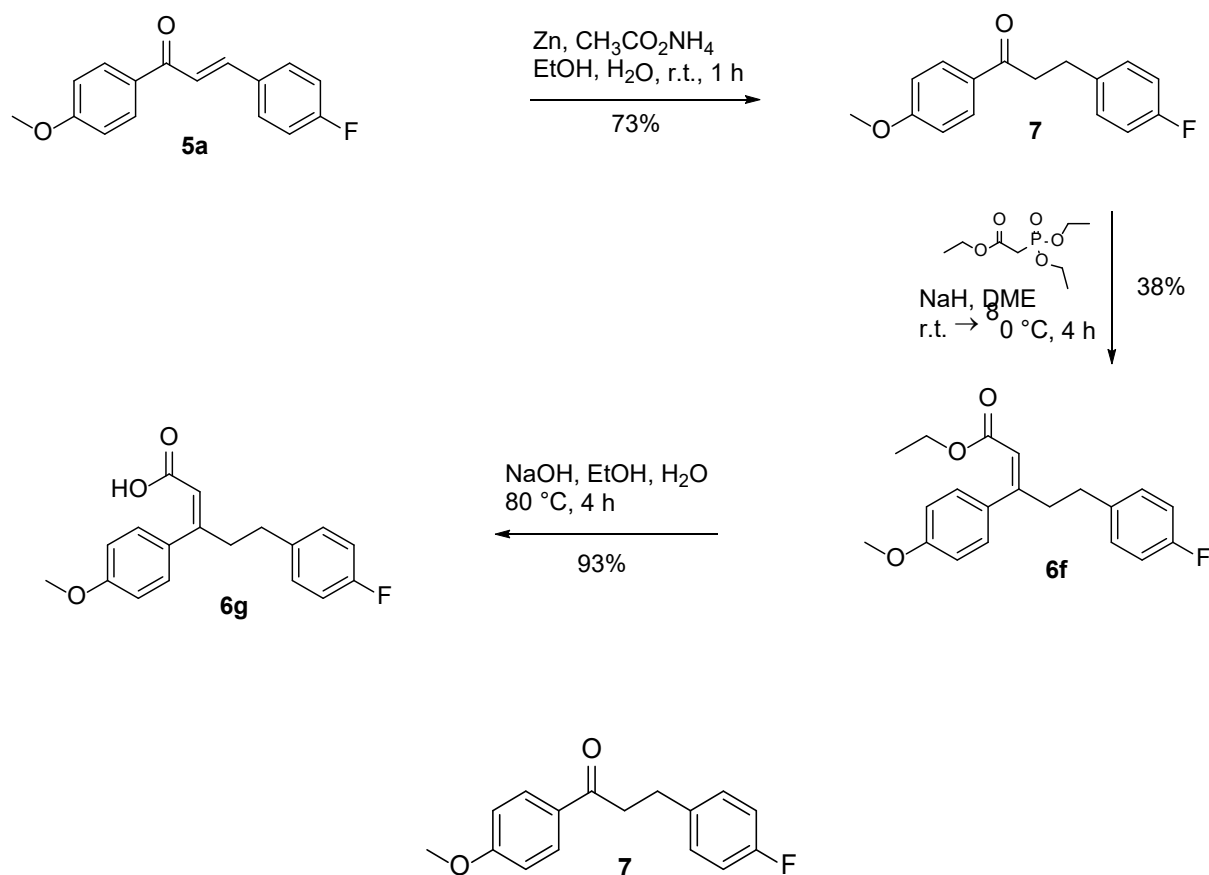
[M+Na]⁺

calc.: 321.0893

ESI-HRMS

found: 321.0897

Scheme 4: Synthesis of 6f and 6g



3-(4-Fluorophenyl)-1-(4-methoxyphenyl)propan-1-one (7)^[13]: To a solution of **5a** (200 mg, 0.78 mmol) in EtOH (50 mL) was added a solution of ammonium acetate (6 g, 78 mmol) in water (8 mL). Then zinc powder (382 mg, 5.85 mmol, 7.5 eq) was put in at room temperature in 5 portions at intervals of 15 min. After completion of the reaction within 70 min (monitored by TLC), the mixture was filtered, the filter cake washed with EtOH (2 x 15 mL), and the combined filtrates concentrated in vacuo. The residue was diluted with water, extracted with ethyl acetate (3 x) and the organic phase was dried (Na_2SO_4) and concentrated in vacuo. Crude product was purified by column chromatography on silica gel using 5% ethyl acetate in pet ether to obtain compound **7** (133 mg, Yield: 73%) as a colourless thick liquid.

^1H NMR (300 MHz, CDCl_3) δ 7.99 – 7.79 (m, 2H), 7.27 – 7.06 (m, 2H), 7.03 – 6.79 (m, 4H), 3.84 (s, 3H), 3.20 (ddd, J = 8.0, 6.9, 0.9 Hz, 2H), 3.01 (t, J = 7.5 Hz, 3H).

^{13}C NMR (126 MHz, CDCl_3) δ 197.33, 163.33, 162.17, 160.23, 136.93, 130.15, 129.81, 129.69, 115.10, 113.65, 55.45, 40.07, 29.49.

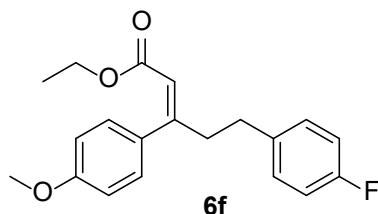
$\text{C}_{16}\text{H}_{15}\text{FO}_2$ (258.2875)

$[\text{M}+\text{Na}]^+$

clac.: 281.0949

ESI-HRMS

found: 281.0948



Ethyl (2Z)-5-(4-fluorophenyl)-3-(4-methoxyphenyl)pent-2-enoate (6f):

Triethylphosphonoacetate (1.15 mL, 5.8 mmol) was added to a suspension of 60% NaH (232 mg, 5.8 mmol) in dimethoxyethane (20 mL) at room temperature and the mixture stirred for 1 h. A solution of compound **7** (500 mg, 1.93 mmol) in dimethoxyethane (5 mL) was slowly added at room temperature and the reaction mixture heated at 80°C for 4 h. After cooling to room temperature, the mixture was diluted with water (100 mL) and extracted with CH₂Cl₂ (3 x). The organic phase was dried (Na₂SO₄) and evaporated in vacuo. The obtained crude material was purified by column chromatography on silica gel using 5 – 10% ethyl acetate in petroleum ether to afford compound **6f** (241 mg, Yield: 38%) as a colourless thick liquid.

¹H NMR (300 MHz, CDCl₃) δ 7.40 (d, *J* = 9.0 Hz, 2H), 7.20 – 7.11 (m, 2H), 6.97 – 6.85 (m, 4H), 6.02 (s, 1H), 4.17 (q, *J* = 7.1 Hz, 2H), 3.83 (s, 3H), 3.40 – 3.26 (m, 2H), 2.75 – 2.62 (m, 2H), 1.29 (t, *J* = 7.1 Hz, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 166.33, 162.17, 160.35, 158.43, 137.14, 132.91, 129.76, 127.94, 116.09, 114.86, 113.97, 59.79, 55.37, 34.55, 32.95, 14.44.

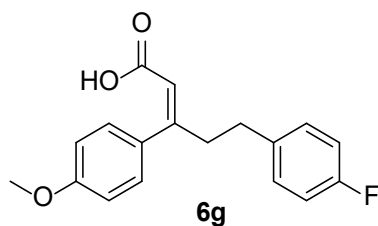
C₂₀H₂₁FO₃ (328.3773)

[M+Na]⁺

calc.: 351.1369

ESI-HRMS

found: 351.1367



(2Z)-5-(4-Fluorophenyl)-3-(4-methoxyphenyl)pent-2-enoic acid (6g): A mixture of **6f** (61 mg, 0.185 mmol) and NaOH (74 mg, 1.85 mmol) in EtOH (4 mL) and water (1 mL) was heated at 80°C for 4 h. The reaction mixture was cooled to room temperature, acidified with 2N HCl, and extracted with ethyl acetate (3 x). The organic phase was dried (Na₂SO₄) and concentrated

in vacuo. The resulting solid was washed with petroleum ether to get the desired compound **6g** (51 mg, Yield: 93%) as a white solid.

¹H NMR (300 MHz, DMSO-*d*₆) δ 12.06 (s, 1H), 7.52 (d, *J* = 8.8 Hz, 2H), 7.22 (dd, *J* = 8.6, 5.7 Hz, 2H), 7.07 (t, *J* = 8.9 Hz, 2H), 6.98 (d, *J* = 8.8 Hz, 2H), 6.00 (s, 1H), 3.80 (s, 3H), 3.37 – 3.26 (m, 2H), 2.69 – 2.57 (m, 2H).

¹³C NMR (126 MHz, DMSO-*d*₆) δ 166.91, 161.32, 159.80, 159.41, 156.88, 137.18, 131.93, 129.64, 127.75, 116.16, 114.60, 113.92, 55.10, 33.90, 31.60.

C₁₈H₁₇FO₃ (300.3242)

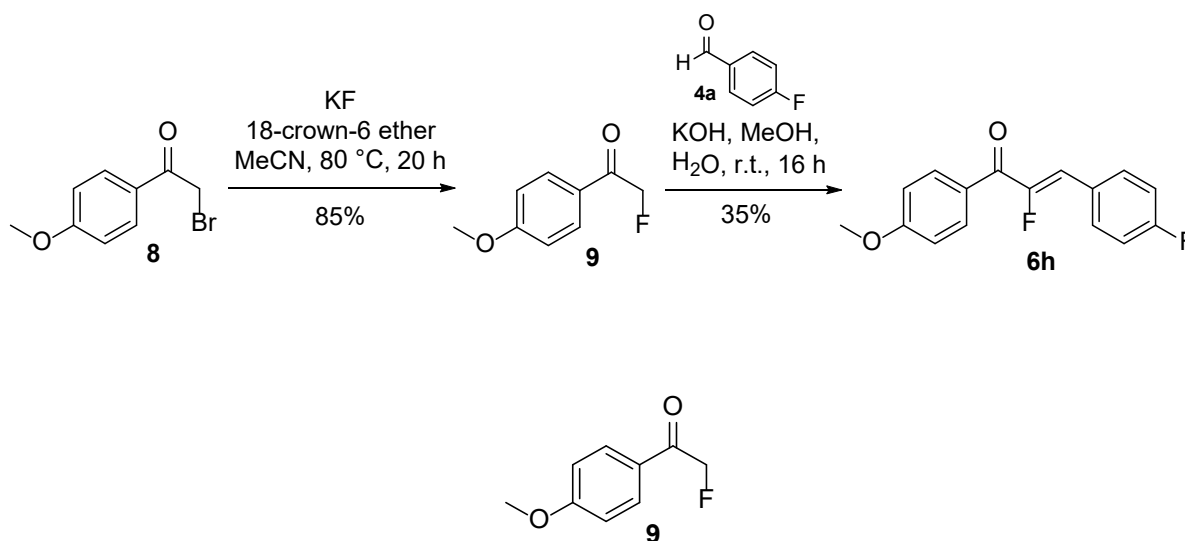
[M+Na]⁺

calc.: 323.1050

ESI-HRMS

found: 323.1054

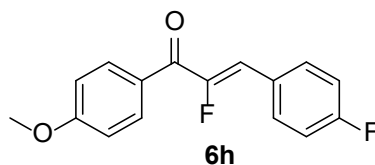
Scheme 5: Synthesis of 6h



2-Fluoro-1-(4-methoxyphenyl)ethan-1-one (9)^[14]: A mixture of 2-bromo-4-methoxyacetophenone (500 mg, 2.18 mmol; labelled as compound **8**), KF (634 mg, 10.9 mmol) and 18-crown-6 ether (115 mg, 0.44 mmol) in CH₃CN (10 mL) was heated at 80°C for 20 h under Argon atmosphere. The reaction mixture was concentrated and the residue was diluted with water, extracted with ethyl acetate (3 x). Combined organic layer was washed with brine, dried (Na₂SO₄), and concentrated under vacuum. Crude product was purified on silica gel column chromatography using 0 – 5% ethyl acetate in pet ether to get the desired compound **9** (310 mg, Yield: 85%) as a white solid.

¹H NMR (300 MHz, CDCl₃) δ 7.93 – 7.78 (m, 2H), 7.03 – 6.82 (m, 2H), 5.56 – 5.37 (m, 2H), 3.86 (s, 3H).

C₉H₉FO₂ (168.1650)



(2Z)-2-Fluoro-3-(4-fluorophenyl)-1-(4-methoxyphenyl)prop-2-en-1-one (6h): 20% aq KOH (0.1 mL) was added to a solution of compound **9** (100 mg, 0.59 mmol) and 4-fluorobenzaldehyde (73.5 mg, 0.59 mmol) in MeOH (2 mL) at room temperature and the mixture was stirred at room temperature for 16 h. Ice cold water was added and the reaction mixture acidified with 2N aq HCl. The resulting precipitate was filtered off, washed with water

(4 x 7 mL) and allowed to dry at the air. The product was purified by column chromatography on silica gel using 5% ethyl acetate in petroleum ether to afford compound **6h** (56 mg, Yield: 35%) as a white solid.

¹H NMR (300 MHz, CDCl₃) δ 7.96 – 7.90 (m, 2H), 7.72 – 7.63 (m, 2H), 7.09 (t, *J* = 8.7 Hz, 2H), 6.96 (d, *J* = 8.9 Hz, 2H), 6.82 (d, *J* = 36.4 Hz, 1H), 3.87 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 185.61, 164.08, 163.53, 162.08, 155.69, 153.51, 132.36, 131.84, 128.52, 127.73, 117.37, 115.93, 113.73, 55.52.

C₁₆H₁₂F₂O₂ (274.2621)

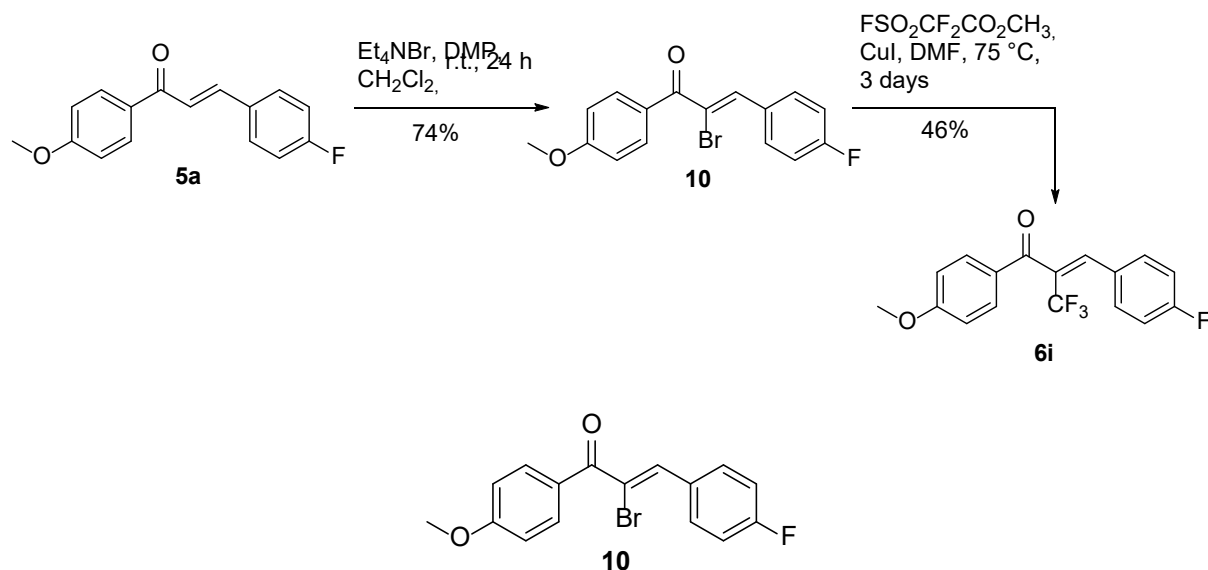
[M+Na]⁺

calc.: 297.0700

ESI-HRMS

found: 297.0698

Scheme 6: Synthesis of 6i



((2Z)-2-Bromo-3-(4-fluorophenyl)-1-(4-methoxyphenyl)prop-2-en-1-one (10): Et_4NBr (984 mg, 4.68 mmol) was added to a stirred suspension of Dess–Martin periodinane (DMP) (1.98 g, 4.68 mmol) in CH_2Cl_2 (8 mL) at room temperature and stirring was continued for 10 min with formation of a pale yellow solution. Compound **5a** (1 g, 3.9 mmol) was added to the mixture in portions at room temperature and stirring was continued at room temperature for 24 h. Then the mixture was diluted with CH_2Cl_2 (50 mL), washed with a sat solution of NaHSO_3 and a sat solution of NaHCO_3 and then with water. The organic phase was dried (Na_2SO_4) and concentrated in vacuo. Purification by column chromatography on silica gel using 5% ethyl acetate in petroleum ether yielded compound **10** (970 mg, Yield: 74%) as a pale-yellow thick liquid.

^1H NMR (300 MHz, CDCl_3) δ 7.96 – 7.89 (m, 2H), 7.25 (s, 1H), 7.18 – 7.09 (m, 2H), 6.91 – 6.82 (m, 4H), 3.83 (s, 3H).

^{13}C NMR (126 MHz, CDCl_3) δ 190.34, 164.49, 163.39, 161.40, 133.91, 132.37, 129.88, 126.09, 115.64, 114.21, 113.81, 55.56.

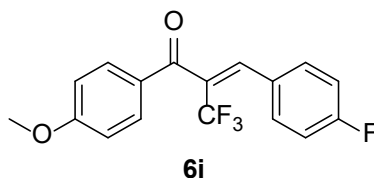
$\text{C}_{16}\text{H}_{12}\text{BrFO}_2$ (335.1677)

$[\text{M}+\text{Na}]^+$

calc.: 356.9886

ESI-HRMS

found: 356.9897



(2Z)-3-(4-Fluorophenyl)-1-(4-methoxyphenyl)-2-(trifluoromethyl)prop-2-en-1-one (6i): A mixture of compound **10** (200 mg, 0.7 mmol), CuI (136 mg, 0.72 mmol) and methyl 2,2-difluoro-2-(fluorosulfonyl) acetate (0.55 mL, 4.35 mmol) in dry DMF (20 mL) was heated at 75°C for 3 days under an argon atmosphere. Then, the reaction mixture was cooled to room temperature, diluted with ether (50 mL), filtered, the filter cake washed with ether and the combined filtrates mixed with water (150 mL) and extracted with ether. The organic phase was washed with brine, dried (Na₂SO₄) and concentrated in vacuo. Purification by column chromatography on silica gel using 10% ethyl acetate in petroleum ether yielded compound **6i** (89 mg, Yield: 46%) as colourless thick liquid.

¹H NMR (300 MHz, CDCl₃) δ 7.85 (d, *J* = 9.0 Hz, 2H), 7.36 (d, *J* = 1.8 Hz, 1H), 7.29 – 7.19 (m, 2H), 6.92 – 6.79 (m, 4H), 3.81 (s, 3H).

¹³C NMR (126 MHz, Chloroform-*d*) δ 190.61, 164.54, 164.29, 162.29, 134.63 (q, *J* = 5.9 Hz), 132.03, 131.46 (d, *J* = 8.6 Hz), 128.33, 128.24 (d, *J* = 3.4 Hz), 123.29, 121.12, 115.85 (d, *J* = 21.9 Hz), 114.12, 55.53.

C₁₇H₁₂F₄O₂ (324.2696)

[M+Na]⁺

calc.: 347.0667

ESI-HRMS

found: 347.0666

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